Further Biphenyl Lignans with a Tetrahydrofuran Moiety from Gymnotheca chinensis

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Four new biphenyl lignans, 1-4, with a tetrahydrofuran moiety were isolated from the MeOH extract of the whole plant of *Gymnotheca chinensis*. Their chemical structures were established by means of spectroscopic analysis, including 1D and 2D NMR spectroscopy and comparison with previously reported data. Compounds 1-4 occur as mixtures of atropisomers as revealed by variable-temperature NMR experiments.

Introduction. – Gymnotheca chinensis DECNE, one of the endemic genera of seed plants in China, is a perennial herb of the family Saururaceae. The whole plants of G. chinensis have long been used as traditional herbal medicine to treat contusions and strains [1]. In our previous investigations on G. chinensis, 15 new lignans belonging to the dibenzocyclooctene, eupomatilone, and eupodienone series were isolated [2]. Eupomatilones are biphenyl lignans originally isolated from the Australian shrub *Eupomatia bennettii*. All members of this family contain a highly oxygenated biphenyl system with a γ -lactone ring or a tetrahydrofuran ring attached to one of the aryl rings, and some of the eupomatilone molecules occur as a mixture of fluctional atropisomers [3]. So far, this kind of degraded lignan was only isolated from E. bennettii and G. chinensis. As a consequence, these compounds have attracted interest of synthetic chemists [4–10]. Our recent phytochemical studies led to the isolation of four further new biphenyl lignans with a tetrahydrofuran moiety, named gymnothelignans P–S (1–4, resp.). The compounds occur as mixtures of atropisomers as revealed by variable-temperature NMR experiments.

Results and Discussion. – A MeOH extract of the whole plant of *G. chinensis* was suspended in H_2O , and successively extracted with petroleum ether and AcOEt. The petroleum ether and AcOEt extracts were combined and then further fractionated by various chromatographic techniques to yield four new biphenyl lignans 1-4 (*Fig. 1*).

Compound **1** was obtained as white powder, with the molecular formula, $C_{22}H_{26}O_7$, deduced from HR-ESI-MS (m/z 401.1571 ($[M-H]^-$), indicating ten degrees of unsaturation. In the NMR spectra (*Tables 1* and 2), several signals were doubled, indicating the presence of two rotational stereoisomers in a *ca.* 1:1 ratio at room temperature. On raising the temperature of a (D_6)DMSO solution of **1**, the ¹H-NMR

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Fig. 1. Structures of compounds 1-4

signals broadened and coalesced, eventually yielding a single signal (Fig. 2), owing to the rapid interconversion of the two conformers at high temperature. The coalescence point for the two sets of signals was observed at *ca.* 100°. The presence of a OCH₂O group was evidenced by the HMBCs (Fig. 3) of OCH₂O to C(4') and C(5'). The correlations of MeO(6') to C(6'), and MeO(3'') to C(3'') positioned the two MeO groups to C(6') and C(3''), respectively. Similarly, a MeO group at C(2) was also assigned. HMBCs H-C(2)/C(5) indicated the connection of C(2) to C(5) via an ether bond. Detailed HMBC analysis suggested that the structure of 1 was very similar to that of gymnothelignan G [2], except for the absence of MeO at C(5''), which was revealed by the ¹H-NMR AMX spin system (δ (H) 6.57 (br. d, J = 8.0, H-C(6'')), 6.82 (d, J = 8.0, H-C(5'')), 6.70 (br. s, H-C(2''))). The orientations of the substituents of the tetrahydrofuran ring were deduced from NOESY correlations (Fig. 4). The crosspeaks H-C(2)/Me(7) and H-C(5)/Me(6) indicated that H-C(2) and Me(7) were cofacial, H-C(5) and Me(6) were on the opposite face. The vicinal coupling constants J(2,3) = 4.9 and J(4,5) = 4.1 were in close agreement with those observed in gymnothelignan G [2]. Gymnothelignan P (1) was thus identified as 5-(4"-hydroxy-6',3''-dimethoxy-4',5'-(methylenedioxy)-1,1'-biphenyl-2'-yl)- 2α -methoxy- $3\beta,4\alpha$ -dimethyltetrahydrofuran¹). The absolute configuration of **1** remains to be determined.

Compound **2** and **3** had the same molecular formula as those of **1** as deduced from HR-ESI-MS. The ¹H- and ¹³C-NMR, and HMBC spectra indicated that **2** and **3** were stereoisomers of **1**. In a NOESY experiment, the cross-peaks H-C(2)/Me(7), H-C(2)/H-C(5), and H-C(3)/H-C(4) were observed in compound **2**, and H-C(2)/H-C(3), H-C(2)/Me(6), H-C(3)/Me(6), and H-C(5)/H-C(4), Me(7)/H-C(4) were observed

¹⁾ For systematic names, see the Exper. Part.

	Table 1. $^{I}H_{-}$	NMR Data (400 MHz) of 1	4 . ð in ppm, <i>J</i> in H	z. Atom numbering as	s indicated in Fig. 1.	
Position	1 ^a)	1 ^b)	1 ^c)	2 ^a)	3 ^a)	4 ^a)
2	4.97 (d, J = 4.9)	4.95 $(d, J = 4.9)$	4.96(d, J = 4.9)	4.55 (s)	4.91 (d, J = 4.4)	5.01 $(d, J = 4.4)$
	4.98 (d, J = 4.9)	$4.96 \ (d, J = 4.9)$		4.55(s)	4.91 (d, J = 4.4)	5.00 (d, J = 4.4)
3	$2.30 - 2.41 \ (m)$	2.25-2.36 (m)	$2.29 - 2.38 \ (m)$	2.28 - 2.38 (m)	$1.83 - 1.89 \ (m)$	2.31–2.43 (<i>m</i>)
	$2.30 - 2.41 \ (m)$	2.25 - 2.36 (m)		2.28 - 2.38 (m)	$1.83 - 1.89 \ (m)$	$2.31 - 2.43 \ (m)$
4	$1.91 - 2.00 \ (m)$	$1.86 - 1.94 \ (m)$	$1.93 - 2.01 \ (m)$	2.10-2.18(m)	$1.75 - 1.83 \ (m)$	$1.93 - 2.02 \ (m)$
	$1.91 - 2.00 \ (m)$	1.86 - 1.94 (m)		2.10-2.18 (m)	1.75 - 1.83 (m)	$1.93 - 2.02 \ (m)$
5	4.58(d, J = 4.1)	$4.42 \ (d, J = 4.0)$	4.47 (d, J = 4.6)	4.52 (d, J = 8.9)	4.99(d, J = 8.5)	4.65 (d, J=3.9)
	4.55(d, J=4.1)	$4.42 \ (d, J = 4.0)$		4.52 (d, J = 8.9)	5.04(d, J = 8.5)	$4.62 \ (d, J=3.9)$
6	$0.61 \ (d, J = 7.2)$	0.53 (d, J = 7.1)	0.60 (d, J = 7.1)	0.67 (d, J = 6.9)	0.66(d, J = 7.5)	$0.61 \ (d, J = 7.1)$
	0.62 (d, J = 7.2)	0.53 (d, J = 7.1)		0.68 (d, J = 6.9)	0.68 (d, J = 7.5)	0.62 (d, J = 7.1)
7	0.85 (d, J = 7.2)	$0.79 \ (d, J = 7.2)$	0.82 (d, J = 7.2)	$0.74 \ (d, J = 7.3)$	0.92 (d, J = 6.6)	0.85 (d, J = 7.2)
	0.86 (d, J = 7.2)	$0.79 \ (d, J = 7.2)$		0.75 (d, J = 7.3)	0.92 (d, J = 6.6)	0.86 (d, J = 7.2)
3,	6.70(s)	6.68(s)	6.68(s)	6.85(s)	6.64(s)	(s) 00(s)
	6.70(s)	6.68(s)		6.85(s)	6.64(s)	(s) 00(s)
2"	6.77 (d, J = 1.9)	$6.66\ (d, J = 1.8)$	6.70 (br. s)	6.74 (br. s)	6.70 (br. s)	6.78 (d, J = 1.8)
	6.75 (d, J = 1.9)	$6.69 \ (d, J = 1.8)$		6.74 (br. s)	6.74 (br. s)	$6.80 \ (d, J = 1.8)$
5''	(6.88 (d, J = 8.0))	$6.80 \ (d, J = 8.0)$	(6.82 (d, J = 8.0))	$6.84 \ (d, J = 8.1)$	6.88 (d, J = 8.0)	6.88 (d, J = 8.0)
	(0.88 (d, J = 8.0))	$6.80 \ (d, J = 8.0)$		6.85 (d, J = 8.1)	6.88 (d, J = 8.0)	6.88 (d, J = 8.0)
6''	$6.61 \ (dd, J = 8.0, 1.9)$	$6.51 \ (dd, J = 8.0, 1.8)$	6.57 (d, J = 8.0)	6.60 (d, J = 8.1)	6.59 (d, J = 8.0)	$6.64 \ (dd, J = 8.0, 1.8)$
	$6.66 \ (dd, J = 8.0, 1.9)$	$6.56 \ (dd, J = 8.0, 1.8)$		6.62 (d, J = 8.1)	6.63 (d, J = 8.0)	$6.69 \ (dd, J = 8.0, 1.8)$
2-MeO	3.25(s)	3.18(s)	3.22(s)	3.38(s)	3.21(s)	3.25(s)
	3.25(s)	3.19(s)		3.38(s)	3.21(s)	3.25(s)
OCH_2O	6.00, 6.01 (2s)	6.00, 6.03 (2s)	5.99, 6.00 (2s)	6.02(s)	6.01(s)	
	6.00, 6.01 (2s)	6.00, 6.03 (2s)		6.02(s)	6.01(s)	
6'-MeO	3.73(s)	3.68(s)	3.69(s)	3.72(s)	3.74(s)	3.56(s)
	3.73(s)	3.69(s)		3.72(s)	3.74(s)	3.56(s)
3''-MeO	3.83(s)	3.71(s)	3.77(s)	3.82(s)	3.83(s)	3.84(s)
	3.85(s)	3.74(s)		3.84(s)	3.84(s)	3.86(s)
4'-MeO						3.87(s)
						3.87(s)
5'-MeO						3.81(s)
						3.81(s)
4″-OH	7.59(s)	8.98 (s)	8.44 (br. s)	7.57 (br. s)	$7.60 (\mathrm{br.}s)$	$7.58 ({\rm br.}s)$
	7.59 (s)	8.98 (s)		7.57 (br. s)	$7.60 ({\rm br.}s)$	7.59 (br. s)
^a) Recorded	in (D_6) acetone, at 25°. ^b) Recorded in (D ₆)DMSO,	at 25°. °) Recorded i	n (D ₆)DMSO, at 100°		

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Table 2. ¹³C-NMR Data (100 MHz, in (D₆)acetone, at 25°) of **1–4**. δ in ppm. Atom numbering as indicated in Fig. 1.

Position	1	2	3	4
2	108.3	111.5	106.2	108.3
	108.2	111.5	106.2	108.3
3	40.3	44.0	42.6	40.2
	40.3	44.0	42.6	40.2
4	44.3	44.4	46.5	44.2
	44.4	44.5	46.8	44.3
5	85.0	84.0	79.5	85.1
	85.0	84.0	79.5	85.1
6	15.1	11.6	16.3	15.2
	15.2	11.6	16.5	15.3
7	9.1	11.2	12.6	9.1
	9.1	11.2	12.6	9.1
1′	128.5	128.4	128.6	128.7
-	128.6	128.5	128.7	128.8
2'	137.9	137.1	136.9	138.9
	137.9	137.1	136.9	138.9
3'	101.3	102.7	102.9	105.9
	101.3	102.7	102.9	105.9
4′	149.3	149.5	150.0	153.6
	149.3	149.5	150.0	153.6
5'	136.8	136.3	134.4	142.1
	136.8	136.3	134.4	142.1
6'	141.9	141.4	141.9	152.2
	141.9	141.4	141.9	152.2
1″	128.2	129.8	128.4	128.6
	128.2	129.8	128.4	128.7
2"	115.3	115.1	115.3	115.3
-	115.5	115.3	115.3	115.4
3″	148.0	147.7	147.8	147.9
	148.0	147.8	147.9	148.0
4''	146.5	146.5	146.4	146.4
	146.5	146.5	146.5	146.5
5″	115.4	114.4	114.3	114.2
	115.9	115.9	115.6	115.8
6″	123.3	123.5	123.5	123.3
	124.9	125.0	124.2	124.9
2-MeO	54.8	54.9	54.5	54.8
	54.9	55.0	54.6	54.8
OCH ₂ O	102.0	102.0	102.0	51.0
	102.0	102.0	102.0	
6′-MeO	60.0	60.0	60.0	61.1
0 11100	60.0	60.0	60.0	61.1
3″-MeO	56.4	56.3	56.3	56.4
s -meo	56.4	56.3	56.3	56.4
4'-MeO	50.7	50.5	50.5	56 3
1 1100				56.3
5′-MeO				60.7
5 1100				60.7
				00.7



Fig. 2. Temperature dependence of the ¹H-NMR signals (400 MHz, in (D_6)DMSO) of the aryl H-atoms of **1**



Fig. 3. Key HMB $(H \rightarrow C)$ correlations of compound 1



Fig. 4. Key NOESY $(H \leftrightarrow H)$ correlations of compound 1

in compound **3**. The vicinal coupling constants $J(2,3) \approx 0$ in **2** and J(2,3) = 4.4 in **3**, and J(4,5) = 8.9 in **2** and J(4,5) = 8.5 in **3** were in close agreement with those observed in gymnothelignans F and H [2]. Thus, gymnothelignan Q (**2**) and gymnothelignan R (**3**) were identified as 2,3-diepi-**1** and 3,4-diepi-**1**, respectively.

Compound **4** was obtained as white powder. The molecular formula was determined as $C_{23}H_{30}O_7$ by analysis of the HR-ESI-MS (m/z 441.1891 ($[M + Na]^+$), indicating nine degrees of unsaturation. The ¹H- and ¹³C-NMR spectra (*Tables 1* and 2) were very similar to those of **1** except for the replacement of the OCH₂O moiety by two MeO groups at C(4') and C(5'). Gymnothelignan S (**4**) was then determined as 5-(4''-hydroxy-4',5',6',3'-tetramethoxy-1,1'-biphenyl-2'-yl)-2 α -methoxy-3 β ,4 α -dimethyl tetra-hydrofuran¹).

In conclusion, we identified four new biphenyl lignans with a tetrahydrofuran moiety, gymnothelignans P-S (1-4, resp.), from *G. chinensis*. These compounds were shown to have the same C-skeleton as eupomatilones, and may be biogenetically derived from eupodienones [2][11]. Besides the eupomatilones, no structurally related compounds were found in plants.

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Experimental Part

General. Column chromatography (CC): silica gel (SiO₂, 300–400 mesh; Qingdao Marine Chemica Co.), MCI CHP-20 gel (75–150 μ m; Mitsubishi), and ODS (40–63 μ m; LiChroprep). TLC: Precoated plates GF 254 (Qingdao Marine Chemical Co.). Semi-prep. HPLC: LC3000 System (Beijing

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ChuangXingTongHeng Science and Technology Co., Ltd.); column, C18 (5 µm, i.d. 10×250 mm; Kromasil). Optical rotations: PerkinElmer-241 polarimeter. UV Spectra: PerkinElmer Lambda 35. IR Spectra: Nicolet-MX-1 spectrometer; $\tilde{\nu}$ in cm⁻¹. NMR Spectra: Bruker Ascend 400 instrument; δ in ppm rel. to Me₄Si as internal standard, J in Hz. HR-ESI-MS: MicrOTOF-Q II mass spectrometer (Bruker, Germany); in m/z. ESI-MS: Micromass Xevo triple-quadrupole mass spectrometer (Waters Corp., USA); in m/z.

Plant Material. The plant was collected from Jinfoshan in Chongqing City, P. R. China, in October 2012, and identified as *G. chinensis* by Prof. *S. R. Yi*, Chongqing Institute of Pharmaceutical Plant. A voucher specimen (20130125) was deposited with the herbarium of the Chengdu Institute of Biology, Chinese Academy of Sciences.

Extraction and Isolation. Dried and powdered whole plants of *G. chinensis* (1.45 kg) were extracted with MeOH at r.t. to give an extract (152 g), which was suspended in H₂O (11) and extracted with petroleum ether (PE) and AcOEt (3×11 , 3 h each) successively. The PE and AcOEt extracts (8 and 19 g, resp.) were combined and then subjected to CC (*MCI* gel (85×100 mm); MeOH/H₂O 90:10). In total, two fractions, *A* and *B*, were obtained based on TLC analysis. *Fr. A* (15 g) was further separated by medium-pressure CC (SiO₂ (49×460 mm); PE/acetone $100:1 \rightarrow 1:1$) to give twelve fractions. *Fr. 7* (800 mg) was purified by CC (*ODS* ($40-63 \mu m, 36 \times 310 mm$); MeOH/H₂O $40:60 \rightarrow 100:0$) to afford three main fractions, *Frs. 7A* – 7*C*. Compound **4** (t_R 24.2 min; 4 mg) was obtained from *Fr. 7A* by semi-prep. HPLC (MeOH/H₂O 66:34; flow rate, 3 ml/min). Compounds **2** (t_R 23.1 min; 8 mg), **3** (t_R 35.4 min; 4 mg), and **1** (t_R 38.3 min, 10 mg) were obtained from *Fr. 7B* by semi-prep. HPLC (MeOH/H₂O 70:30; flow rate, 3 ml/min).

Gymnothelignan P (=2-*Methoxy*-4-{4-*methoxy*-6-[($2R^*, 3R^*, 4R^*, 5R^*$)-2,3,4,5-*tetrahydro*-5-*methoxy*-3,4-*dimethylfuran*-2-*yl*]-1,3-*benzodioxo*l-5-*yl*]*phenol*; **1**). White powder. [a]₂₀²⁰ = -90 (c = 0.04, MeOH). UV (MeOH): 267 (3.69). IR (KBr): 3421, 2935, 1596, 1518, 1487, 1463, 1402, 1253, 1195, 1137, 1094, 1010. ¹H- and ¹³C-NMR: *Tables 1* and 2, resp. ESI-MS: 401 ([M - H]⁻), 425 ([M + Na]⁺). HR-ESI-MS: 401.1571 ([M - H]⁻, C₂₂H₂₅O₇⁻; calc. 401.1600).

Gymnothelignan Q (=2-*Methoxy*-4-[4-*methoxy*-6-[(2*R**,3*R**,4*S**,5*S**)-2,3,4,5-*tetrahydro*-5-*methoxy*-3,4-*dimethylfuran*-2-*yl*]-1,3-*benzodioxo*l-5-*yl*]*phenol*; **2**). White powder. $[\alpha]_D^{20} = -64$ (c = 0.048, MeOH). UV (MeOH): 267 (3.49). IR (KBr): 3276, 2970, 1598, 1518, 1477, 1390, 1233, 1212, 1082, 1034, 916. ¹H- and ¹³C-NMR: *Tables 1* and 2, resp. ESI-MS: 401 ([M - H]⁻), 425 ([M + Na]⁺). HR-ESI-MS: 401.1568 ([M - H]⁻, C₂₂H₂₅O₇; calc. 401.1600).

Gymnothelignan R (=2-*Methoxy*-4-{4-*methoxy*-6-[($2R^*$, $3S^*$, $4S^*$, $5R^*$)-2,3,4,5-*tetrahydro*-5-*methoxy*-3,4-*dimethyltetrahydrofuran*-2-*yl*]-1,3-*benzodioxo*l-5-*yl*]*phenol*; **3**). White powder. [a]²⁰_D = -97 (c = 0.041, MeOH). UV (MeOH): 267 (3.60). IR (KBr): 2961, 1616, 1518, 1476, 1374, 1269, 1081, 936. ¹H- and ¹³C-NMR: *Tables 1* and 2, resp. ESI-MS: 401 ([M - H]⁻), 425 ([M + Na]⁺). HR-ESI-MS: 401.1604 ([M - H]⁻, C₂₂H₂₅O₇⁻; calc. 401.1600).

Gymnothelignan S (=2',3,3',4'-Tetramethoxy-6'-[($2R^*$,3 R^* ,4 R^* ,5 R^*)-2,3,4,5-tetrahydro-5-methoxy-3,4-dimethylfuran-2-yl]-1,1'-biphenyl-4-ol; **4**). White powder. [a]_D²⁰ = -17 (c = 0.036, MeOH). UV (MeOH): 266 (3.43). IR (KBr): 3421, 2935, 1597, 1519, 1487, 1463, 1253, 1138, 1094, 1010. ¹H- and ¹³C-NMR: *Tables 1* and 2, resp. ESI-MS: 417 ([M - H]⁻), 441 ([M + Na]⁺). HR-ESI-MS: 441.1884 ([M + Na]⁺, C₂₃H₃₀NaO₇⁺; calc. 441.1889).

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